

Drip Irrigation in California Strawberry Nurseries to Reduce the Incidence of *Colletotrichum acutatum* in Fruit Production

Oleg Daugovish

University of California Cooperative Extension, Ventura County, 669 County Square Drive, Suite 100, Ventura, CA 93003

Mark Bolda

University of California Cooperative Extension, Santa Cruz County, 1432 Freedom Boulevard, Watsonville, CA 95076

Sukhwinder Kaur

Department of Plant Pathology, University of California, Davis, CA 95616-8680

Maren J. Mochizuki

University of California Cooperative Extension, Ventura County, 669 County Square Drive, Suite 100, Ventura, CA 93003

Daniel Marcum

University of California Cooperative Extension, P.O. Box 9, 44218 A Street, McArthur, CA 96056

Lynn Epstein¹

Department of Plant Pathology, University of California, Davis, CA 95616-8680

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Abstract. Strawberry anthracnose caused by *Colletotrichum acutatum* is often asymptomatic on plants in sprinkler-irrigated nurseries but destructive after transplantation into fruiting fields. This study evaluated the impact of strawberry nursery sprinkler and drip irrigation in the presence or absence of *C. acutatum* on post-transplantation plant growth, mortality, and fruit yield in fruiting fields in California. In a 2005 nursery at Tulelake, CA, dip infestation of mother plants with *C. acutatum* reduced early-season mother plant canopy size by 21% and delayed runner production in mother plants but otherwise had no obvious disease symptoms. In comparison with sprinkler-irrigated treatments in the nursery, drip irrigation of infested nursery plots reduced plant losses in fruit production fields by 86% at Watsonville, CA, in 2005 and 50% and 75% at Oxnard, CA, in 2005 and 2008. Transplants from infested nursery treatments had a 33% to 60% smaller canopy and 11% to 42% lower yield than transplants from uninfested nursery treatments. However, transplant canopy size and yield from the infested and then drip-irrigated nursery treatment were similar to the sprinkler-irrigated, non-infested nursery treatment. Quantitative polymerase chain reaction measurements of *C. acutatum* in crown tissue of fruiting field plants seven weeks after transplanting showed significantly more *C. acutatum* ($\approx 11\times$) in their crowns in sprinkler-irrigated than drip-irrigated, infested nursery transplant treatments. During the course of fruit production, the amount of *C. acutatum* in crown tissue increased in all treatments. However, at the end of the fruit season, there was still significantly more ($\approx 8\times$) *C. acutatum* in the crowns of the plants produced by sprinkler irrigation than by drip irrigation in the nursery. These data suggest that if *C. acutatum* is present in the nursery, drip irrigation can reduce subsequent plant stunting and yield losses in strawberry production fields.

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¹To whom reprint requests should be addressed; e-mail lepstein@ucdavis.edu.

Each year, California strawberry nurseries transplant “mother” plants into 1,000 ha to produce over one billion runner plants (daughters) for either out-of-state sale or for 15,620 ha of California fruit production, valued at \approx \$1.8 billion (Calif. Dept. of Food and Ag., 2010). Nearly all (greater than 95%) transplants originate from individual axenic tissue-culture plants that are used to produce daughters in hanging pots

in a greenhouse followed by a generation in pathogen-free soil in a screenhouse and finally a series of field-grown generations in conditions that promote runner plant production. In early generations of vegetative propagation, mothers produce ≈ 100 daughters each, which declines to ≈ 30 daughters each in the final multiplication in a high-elevation (greater than 1000 m) nursery. At high elevation, the shortening days in August and September stimulate flower bud initiation and facilitate dormancy, and the cool fall temperatures stimulate vigor for early transplantation into coastal fruit production areas in California (Strand, 2008).

Colletotrichum acutatum Simmonds causes anthracnose of strawberry in California and is an internationally regulated pest (EPPO, 2004). The disease is generally asymptomatic in the nursery, but after planting infected transplants into fruiting fields, the plants can be stunted and die. Also, fruit can develop lesions and be unmarketable (Koike and Browne, 2008; Smith, 2008). Consequently, although strawberry anthracnose is only an intermittent problem in California fruiting fields, it is of constant concern in nurseries because nursery plants with the pathogen cause disease outbreaks in fruiting fields for which fruit growers demand compensation from nurseries.

Hot water dips of mother plants are sometimes used in early nursery generations to control nematodes and *C. acutatum*, but the strategy is not used in the high-elevation generation destined for fruiting fields because hot water inhibits vigor (Koike and Browne, 2008) and has the potential to spread *Xanthomonas fragariae*, an internationally regulated pest (Clover et al., 2010). Fungicide dips or water-wash treatments may reduce severity of *C. acutatum* but do not completely eliminate the pathogen (Daugovish et al., 2008; Johnson et al., 2006; Paredes and Munoz, 2002). Because almost all nurseries use sprinkler irrigation, which is conducive for dissemination and infection of *Colletotrichum* species, nurseries routinely spend over \$1200 per ha for fungicides as a preventive treatment for *C. acutatum*.

Anthracnose in strawberry caused by *C. acutatum* is difficult to control, partly because some isolates have a wide host range on crops and weeds (Freeman et al., 2001; Peres et al., 2005) and the pathogen can survive in soil on crop debris (Feil et al., 2003; Urena-Padilla et al., 2001). Because *C. acutatum* conidia are transported by splashing water (Ntahimpera et al., 1997), and water on the leaf surface provides a conducive environment for germination and infection, we postulated that drip irrigation in the nursery would reduce the impact of *C. acutatum* in fruiting fields. The objective of this study was to evaluate the impact of irrigation method and the presence of pathogen pressure in a high-elevation strawberry nursery on subsequent performance of transplants in fruiting fields in California.

Materials and Methods

Growth of nursery transplants. In both years, transplants were produced in Manteca, CA, harvested without leaves, and stored at

-2 °C before use as mother plants in an experimental high-elevation nursery at the Intermountain Research and Extension Center (IREC) at Tulelake, CA (1242 m elevation). In 2005, before the experiment was started, the Tulebasin mucky silty clay loam soil was fumigated with 330 kg·ha⁻¹ 67:33 methyl bromide/chloropicrin on 5 Apr. Four replicated nursery treatments were used in each year: 1) mother plants dip-infested with *C. acutatum* and sprinkler-irrigated; 2) mother plants dip-infested with *C. acutatum* and drip-irrigated; 3) non-infested and sprinkler-irrigated; and 4) non-infested and drip irrigated. Mother plants were thawed, infested when indicated by a 5-min dip of the roots and crowns in a suspension of 2.5×10^5 *C. acutatum* conidia per milliliter, and immediately planted into replicated plots. In 2005, four, 3.4 m × 9.2-m plots per replicate were planted with cv. Ventana on 26 Apr. 2005 with 2.3×10^4 plants/ha (≈ 0.9 m between rows and 0.5-m plant between plants within rows). Four rotary sprinklers and high-flow drip tape with 10-cm distance between emitters were spaced 30 cm apart for the sprinkler- and drip-irrigated plots, respectively. In 2008, 'Camarosa' plants were planted at IREC on 4 Apr. in 2008 at a density of 9×10^4 plants/ha (35-cm plant spacing) in 1.4-m diameter × 25-cm deep bins containing non-fumigated Fordney loamy fine sand soil obtained from uncultivated land adjacent to a commercial nursery near Macdoel, CA; the soil type was selected because it is typical of the high-elevation California strawberry nursery soils. Three randomly assigned replicates per treatment were irrigated by either drip tape in a spiral circle or a central 360° sprinkler on a 1.5-m post. Plots in both years were spaced with 10 m between plots to avoid water movement between plots; in 2005 and 2008, there was barley and gravel between the plots, respectively. Irrigation by both drip and sprinkler was scheduled to maintain conditions of adequate soil moisture at the surface to promote rooting of daughter plants and maximum plant growth. Drip irrigation treatments received only drip irrigation throughout the entire season. At harvest, daughter plants were dug by hand, washed to remove soil, and crowns were sorted. After selection of daughters with crowns that were 7.2 to 10.9 mm in diameter, the plants were cooled and shipped overnight to production fields. In 2005, we conducted an additional trial designed as a completely randomized design in a sprinkler-irrigated plot with 51 blocks with each block having one infested and one non-infested mother plant. These plants were used to measure the impact of infestation early in the season on growth of mother plants and on production of runners (stolons that produce daughter plants).

Plant performance in production fields. In the summers of 2005 and 2008, production fields at Watsonville and Oxnard were flat-fumigated with 330 kg·ha⁻¹ of 67:33 methyl bromide:chloropicrin. Afterward, beds were constructed and covered with black low-density polyethylene mulch 0.15 mm thick.

Two drip lines 8-cm deep irrigated four rows of plants at Oxnard and a single drip line irrigated two rows of plants at Watsonville. The plastic was mechanically slit to permit placement of transplants and overhead sprinklers were used to facilitate establishment in the first six weeks after planting (Strand, 2008). We planted into a randomized complete block design with 10 replicates of each of the four nursery treatments, each with eight plants on 8 Oct. 2005 at both locations and 12 replicates each with eight plants on 3 Oct. 2008 at Oxnard. Each treatment was separated by two rows of commercial 'Ventana' strawberry to prevent cross-contamination between plots.

Six and seven weeks after transplanting in 2005 and 2008, respectively, we counted the total number of dead and undersized plants that would be replanted commercially. Canopy area of plants surviving early mortality was calculated using the formula for an ellipse (plant length/2 × plant width/2 × π).

To adjust for replicates that would be replanted as a result of plant mortality or undersize, four acceptably sized surviving plants per replicate were arbitrarily selected on 15 Nov. 2005 and 19 Nov. 2008 for fruit yield evaluation; the other plants in the plot were removed. The weight of marketable and unmarketable fruit yield was recorded for the four plants per plot for the typical harvest period for each region (Jan. to June 2006 and 2009 at Oxnard and Mar. to Sept. 2006 at Watsonville). As a result of early mortality at Oxnard in 2005, four of the sprinkler/infested plots had less than four plants remaining for yield measurements; consequently, six replications were analyzed for this treatment.

Detection and quantification of *Colletotrichum acutatum* in the daughter plants in the nursery and in the fruiting field. At the indicated sampling times, plants were hand-dug, the soil was removed, the leaves were discarded, and infection by *C. acutatum* was assessed by either bioassay or when indicated in 2009, quantitative real-time polymerase chain reaction (qPCR). Plants were bioassayed for *C. acutatum* using a freezer technique (Mertely and Legard, 2004). Each trimmed plant, with the crown and roots, was washed, frozen at -20 °C for 16 h to kill the plant tissue, surface-disinfested in 0.62% sodium hypochlorite for 2 min, and incubated for 8 to 10 d at 28 °C on moist sterile towels in a re-sealable plastic bag. Crowns and roots were examined under a dissecting microscope for salmon-colored acervuli and putative positives were confirmed with a compound microscope by observation of the expected size and shape of *C. acutatum* conidia.

For qPCR, DNA was quantified using purified *C. acutatum* DNA as an external standard; DNA was purified from the culture used to infest the mother plants as described in Bhat and Browne (2010). The purity and concentration of the standard DNA was determined spectrophotometrically. DNA was isolated from strawberry crowns using a protocol downsized from the one described by

Brandfass and Karlovsky (2008). After plants from the field were washed in running water, the crown was frozen and lyophilized, the outer bark was scraped off, and the crown tissue was ground in liquid N₂ with a mortar and pestle. Forty milligrams of powder was placed in a 2-mL tube with two 4-mm diameter glass beads, 20 mg 106-μm diameter glass beads, and 800 μL of Brandfass and Karlovsky (2008) extraction buffer with polyvinylpyrrolidone 40 instead of polyvinylpolypyrrolidone. Vials were shaken in a FastPrep system (Bio-Spec, Bartlesville, OK) for four cycles of 30 s each. Then, samples were incubated at 42 °C for 10 min, vortexed, and incubated at 65 °C for 10 min with mixing every 3 min. After addition of 640 μL chloroform:isoamyl alcohol (24:1), samples were vortexed 15 s, incubated on ice for 10 min, and centrifuged at 5000 g for 10 min. After 480 μL of the supernatant was transferred to a 1.5-mL tube containing 194 μL 30% (w/v) polyethylene glycol (PEG 6000) and 100 μL 5 M NaCl, the tube was mixed and centrifuged for 15 min at 15,000 g. After the supernatant was discarded, the pellet was washed with 1 mL 70% (v/v) ethanol and centrifuged for 30 s. After removal of the ethanol, the pellet was dried, dissolved in 200 μL 10 mM Tris-HCl (pH 8) containing 1 mM EDTA (TE), and stored at 4 °C.

The following primers and probe (from 5' to 3') were tested by Garrido et al. (2009): ACUT-F1, CGGAGGAAACCAACTCTATTTACA; ACUT-R1, CCAGAACCAAGAATCCGTTG; and 5'Fam-3'Tamra ACUT-PB probe, CGTCTCTTGTGAGTGACACAA GCAAATAATTTAAA. TaqMan® assays (final volume 25 μL) contained 1× Taqman Universal Master mix AB-0055 (Applied Biosystems, Foster City, CA) with ROX, 300 nM of each primer, 100 nM probe, and 5 μL template DNA extract. To assure that PCR inhibitors were removed, both 1:100 and 1:1000 dilutions of template DNA were used. Negative controls without plant extract were included in every run. PCR reactions were conducted at 50 °C for 2 min, 95 °C for 10 min, and 45 cycles of 95 °C for 15 s and 60 °C for 1 min. PCR was performed in a 7500 Fast Real-Time PCR System (Applied Biosystems). The sample cycle threshold values in comparison with the standard were used to calculate the quantity of *C. acutatum* DNA. The absence of PCR inhibitors in the crown extract was assured by the use of the two dilutions of sample DNA and by spiking selected samples with known quantities of standard DNA. *C. acutatum* in roots was not quantified because there were too many PCR inhibitors as indicated by multiple dilutions of sample DNA and by spiking selected samples with known quantities of standard DNA (data not shown).

Statistical analysis. Except when indicated otherwise, data were analyzed using a general linear mixed model (Version 9.2; SAS Systems, Cary, NC). The model assumption for homogenous variance was checked by the Levene test. The overall error rate for multiple comparisons was controlled by Tukey-Kramer honestly significant difference. To address potential autocorrelation for plant size (measured on the same plants on multiple sampling dates),

repeated-measures analysis of variance was used. For this analysis, we selected the compound symmetry and heterogeneous compound symmetry covariance models for Watsonville and Oxnard, respectively, based on measures of relative fit of competing covariance models.

Results and Discussion

Both ‘Camarosa’ and ‘Ventana’ are University of California short-day strawberry varieties. Starting in 2002, ‘Camarosa’ was largely replaced by ‘Ventana’. We used ‘Camarosa’ in the second experiment because it is considered more susceptible to *C. acutatum* (Seijo et al., 2008) and is a prolific producer of daughter plants in California nurseries.

Growth of the nursery transplants. In the nursery in 2005, mother plants that were dip-infested with *C. acutatum* and then grown under sprinkler irrigation were largely asymptomatic; only one runner lesion was observed in 2005. However, plants that were infested with *C. acutatum* were significantly smaller ($P < 0.001$) with an average of a 21% reduction in area in the first 12 weeks (Fig. 1). The infested plants also had significantly ($P < 0.001$) fewer runners in the first 12 weeks with 39% fewer runners than non-infested controls. An analysis of the slope indicated that infested mothers had a significantly ($P = 0.0015$) lower growth rate than the non-infested controls; control mothers increased an average of $49 \pm 2 \text{ cm}^2$ per day, whereas the infested plants increased $40 \pm 2 \text{ cm}^2$ per day. Plant area and the number of runners were significantly correlated ($P < 0.001$, $r = 0.72$ and 0.70 for infested and non-infested, respectively). Infested mothers also had a significantly ($P < 0.0001$) lower rate of runner production than the non-infested controls; control mothers produced an average of 3.2 ± 0.1 new runners per week, whereas the infested plants added 2.1 ± 0.1 runners per week. These data indicate that although the disease is asymptomatic in the nursery, infection can have a negative impact on nursery productivity. Other than a reduction in size and runner production, no other symptoms of *C. acutatum* infection were observed in nursery daughter plants at IREC in either 2005 or 2008. Other researchers (Freeman et al., 2001; Leandro et al., 2001) have observed asymptomatic infections. No adverse impacts of drip irrigation in the nursery on plant production were observed.

In 2005 there were no significant differences in yield of daughter plants between drip and sprinkler irrigation (62.4 and 76.4 m^2 , respectively, $P = 0.08$) (data not shown). In 2008, the infested treatments had 49% fewer daughters; however, the reduced numbers of daughters were specifically in the small-sized category, whereas production of large- and medium-sized daughters was similar in all treatments (Table 1). In commercial practice, in years in which there is a shortage of plants, small plants are used or in some cases harvested later.

Plant growth in fruiting fields. In the fruiting fields, plants from the infested treatments in the nursery generally grew more slowly than plants from the non-infested treatments (Fig. 2). At Watsonville in 2005, daughter plants from infested, sprinkler-irrigated mothers had a 60% smaller canopy size in November and a 66% smaller canopy size in December than plants from infested nursery mothers produced under drip (Fig. 2A). In plants from the non-infested mothers, there was no difference in canopy size between irrigation methods. At Oxnard in 2005–2006 at all sampling dates, plants from drip treatments were similar regardless of whether the mothers were infested (Fig. 2B). Overall, by Jan. 2006, the canopy of plants from infested mothers was 33% smaller than those from non-infested, suggesting the pathogen slowed plant growth. Plants measured in Nov. 2008 at Oxnard showed a similar pattern; the canopy area of plants from infested mothers was 50% less than those from non-infested mothers (Fig. 2C). Although plant canopies developed differently in different years and locations, drip irrigation tended to minimize the negative effects of nursery infestation on early-season canopy size in the fruiting field. The differences in canopy development between production areas and years was not unexpected; fall fertilization, in-field chilling, and harvest date can affect plant growth

and fruit production between locations and years (Galletta and Bringham, 1990).

Plant losses resulting from mortality and poor growth in fruiting fields. Six weeks after transplanting, daughter plants from infested mothers grown with sprinkler irrigation had sevenfold greater mortality at Watsonville in 2005 and two- and fourfold at Oxnard in 2005 and 2008, respectively, compared with daughters from infested mothers produced in drip irrigation (Table 2). In 2005, survival of daughter plants from infested mothers grown with drip irrigation did not differ from the non-infested treatments. In 2008 at Oxnard, in addition to dead plants, plants with diameters less than 7 cm were considered undersized by grower standards and were also removed because growers would typically replant them; because we could cull our undersized plants, none of these plants was replanted in our trial. The four treatments were separated into three replant groups: daughters of infested mothers that were sprinkler-irrigated required significantly more replants, daughters of infested mothers that were drip-irrigated required significantly fewer replants, and daughters of non-infested mothers required no replants (Table 2). In the production field, replants are not always readily available, add to production costs, and usually lag in the fruiting cycle, thereby producing less profit (Larson et al., 2009). Our results suggest that

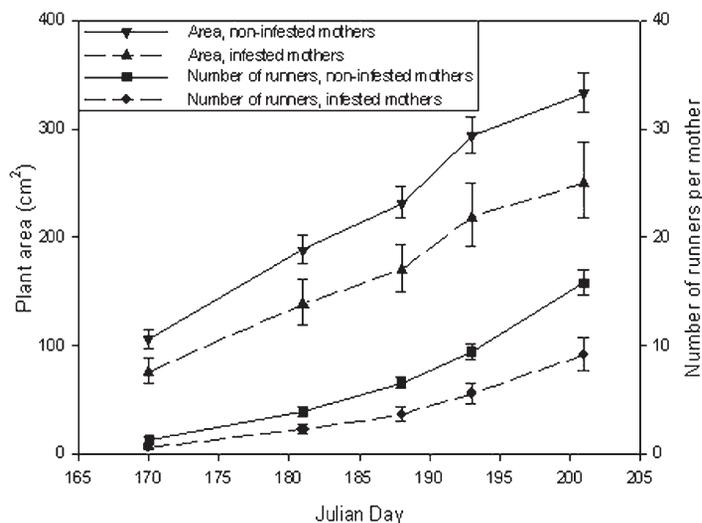


Fig. 1. Effect of *Colletotrichum acutatum* on growth and runner production of ‘Ventana’ strawberry plants in a high-elevation nursery planting with sprinkler irrigation. Data were log-transformed to meet the assumption of additivity for multivariate analysis of variance. Back-transformed means and 95% confidence intervals are shown. $n = 51$.

Table 1. Effect of infestation of mother plant roots with *Colletotrichum acutatum* and sprinkler vs. drip irrigation in the nursery on yield of ‘Camarosa’ daughter plants in the nursery in 2008.^z

Treatment of mothers with <i>C. acutatum</i> at planting	Irrigation during daughter production	Daughters (no. per m ²) ^y			
		Total	Large	Medium	Small
Infested	Sprinkler	167 b	22 a	62 a	83 b
Infested	Drip	147 b	22 a	37 a	88 b
Not infested	Sprinkler	248 a	31 a	77 a	140 ab
Not infested	Drip	263 a	28 a	100 a	145 a

^zAt harvest on 1 Oct. 2008, daughters were classified into three size groups based on crown diameter: large, 11.0 to 17.6 mm; medium, 7.2 to 10.9 mm; and small, 2.5 to 6.9 mm.

^yData met the assumption of homogeneity of variance (Levene $P > 0.05$). Within a column, means followed by the same letter were not significantly different by Tukey’s honestly significant difference, $\alpha = 0.05$.

plants from infested mothers grown with drip irrigation instead of sprinkler in the nursery are less likely to die or be undersized and thus require replanting.

Fruit yields in production fields. A two-way analysis of variance of 2008 yields indicated no significant interaction of the two factors ($P = 0.36$) but a significant effect of both infestation of nursery plants ($P < 0.0001$) with *C. acutatum* and irrigation method ($P = 0.048$). Yields of plants from the non-infested drip or sprinkler-irrigated nursery mothers were similar and generally greater than yields from plants from infested mothers (Table 3). However, plants from infested mothers at Watsonville in 2006 and at Oxnard in 2009 that were drip-irrigated had fruit yields similar to the non-infested sprinkler treatment, consistent with drip irrigation decreasing the negative impact of *C. acutatum* on plant growth in the nursery and in production fields. Interestingly, even after undersized plants were removed, there was a highly significant slope ($P = 0.0001$) of a linear regression of fruit yield on plant area at 5.4 weeks ($r^2 = 0.46$), i.e., plant area at 5.4 weeks after planting predicted almost half of the variance of total cumulative yield (data not shown).

Colletotrichum acutatum infection in daughter plants in the nursery and after fruit production. We used two assays to monitor *C. acutatum* in the daughters: a bioassay in which the fungus is scored as present or absent (Table 4) and a qPCR assay with Taqman probes and *C. acutatum* DNA as an external standard (Tables 4 and 5). At the end of nursery production, there was very low infection (0% to 2%) in the non-infested treatments and high incidence (78% to 82%) in the infested treatments. By bioassay, on a whole-plant basis, the percentage incidence in the infested and either sprinkler- or drip-irrigated was indistinguishable. However, the bioassay indicated that significantly more of the daughters from the infested and sprinkler-irrigated were infected in both the crown and the roots than in the drip-irrigated. At seven weeks after transplanting when the plots were thinned to four plants per replicate, the culled plants were arbitrarily assigned for analysis by either bioassay or qPCR. In terms of percentage infection, results by bioassay and qPCR were indistinguishable at seven weeks after transplanting. However, qPCR expressed as nanograms *C. acutatum* DNA per gram dry weight crown tissue clearly demonstrated that seven weeks after planting, plants that were produced in infested plots with drip irrigation had significantly ($\alpha = 0.05$) less *C. acutatum* in their crowns than plants that were produced in sprinkler-irrigated plots (Table 5). Daughters from infested mothers grown with sprinkler irrigation had significantly more ($\approx 11\times$) *C. acutatum* than daughters from drip-irrigated plots. During the course of fruit production, the amount of *C. acutatum* in crown tissue increased in all treatments. Nonetheless, at the end of the fruit season, there was still significantly more ($\approx 8\times$) *C. acutatum* in the crown tissue of the daugh-

ters that had been produced under sprinkler than under drip in the nursery.

Implications of the research for disease control. There is much anecdotal evidence that asymptomatic but infected nursery transplants are an economically important source of *C. acutatum* in strawberry fruiting fields (Howard et al., 1992; Smith, 2008; Urena-Padilla et al., 2001) and that control of *Colletotrichum* spp. in the nursery is critical

for disease control in fruiting fields (McInnes et al., 1992). *C. acutatum* can grow as an endophyte in strawberry plants (Freeman et al., 2001), conidiate on symptomless leaves (Leandro et al., 2001) and, similar to rain splash, be dispersed by sprinkler irrigation, which also provides a conducive environment for germination and infection of *Colletotrichum* spp. conidia (Madden, 1992; Madden and Wilson, 1997). We provide empirical data

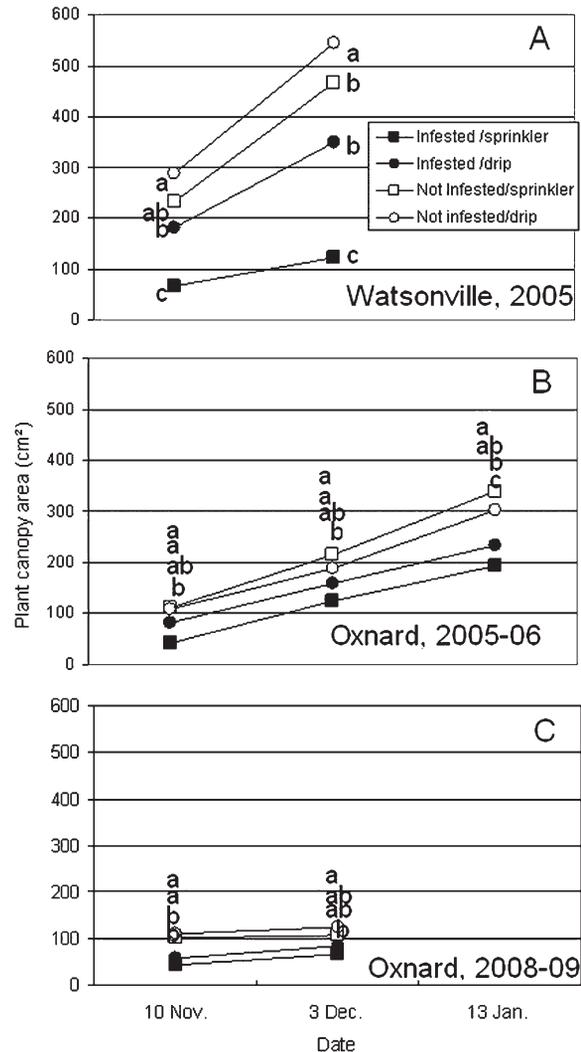


Fig. 2. Effect of nursery treatment on canopy area of surviving strawberry daughter plants before fruit production. Within a location/year, means within a sampling date followed by the same letter are not significantly different ($\alpha = 0.05$; Tukey-Kramer honestly significant difference).

Table 2. Effect of nursery production treatment on either mortality or severe stunting of strawberry daughter plants in fruit production fields.^a

Nursery treatment	Dead or undersized (%) ^b		
	Watsonville 5 Nov. 2005	Oxnard 21 Oct. 2005	Oxnard 24 Oct. 2008
Infested/sprinkler	29 a	40 a	24 a
Infested/drip	4 b	20 b	6 b
Not infested/sprinkler	1 b	15 b	0 c
Not infested/drip	2 b	14 b	0 c

^aMother plants were dip-infested with *Colletotrichum acutatum* and planted in a nursery at Tulelake, CA, in April and then sprinkler- or drip-irrigated until daughters were harvested and planted in the production fields in October.

^bMortality was recorded for eight plants per replicate six weeks after transplanting; there were 10 and 12 replicates in 2005 and 2008, respectively. Undersized plants were less than 7 cm in diameter. Within a column, means followed by the same letter were not significantly different by Fisher's exact test with a Bonferroni adjustment for six pairwise comparisons ($P = 0.0083$ for $\alpha = 0.05$).

that in the presence of *C. acutatum* in the nursery, drip rather than sprinkler irrigation provides a less conducive environment for *C. acutatum* and ultimately results in daughter plants with less fungal growth. Previously, Coelho et al. (2008) showed that in the presence of *C. acutatum* in fruiting fields, there were significantly fewer blighted flowers and rotten fruit in plots with drip rather than overhead irrigation. Although rainfall is critical for *Colletotrichum* spp. in natural environments, summer rainfall in Tulelake

probably did not affect this experiment because rain provided less than 2% of ETo (Calif. Irrigation Management Information System, 1982) in both years in the 3-month period from July to September when most growth in the nursery occurs.

Real-time PCR is increasingly used for both detection and quantification of fungi in plant tissue (Debode et al., 2009; Garrido et al., 2009). Previously, Garrido et al. (2009) used qPCR to quantify *C. acutatum* in artificially inoculated crowns. Because strawberry

transplants in California are transported without leaves, *C. acutatum* DNA in the crowns provides a good proxy for the extent of internal inoculum. Our data provide empirical evidence that strawberry transplants with a pre-existing *C. acutatum* infection are significantly more likely to die early in the production season, be stunted, and/or to have reduced fruit yield. Our data further indicate that in the presence of *C. acutatum* in nursery soil, drip irrigation can significantly decrease the severity of the infection in nursery transplants and thereby have a positive effect on survival, plant growth, and fruit yield.

Although the California strawberry nursery industry predominantly uses sprinkler irrigation, drip is being adopted; one grower uses a combination of drip and sprinklers, and two others use drip irrigation in tunnels, which preclude use of sprinklers. Although material and labor costs for drip irrigation are greater than for sprinkler irrigation, conversion to drip may have compensating savings with reduced water use and less need for irrigation runoff management.

Table 3. Effect of nursery production treatment on post-transplantation strawberry fruit yield.

Nursery treatment	Total marketable yield per plant (g) ^z		
	Watsonville	Oxnard	
	2006	2006	2009
Infested/sprinkler	376 b	403 b	612 c
Infested/drip	473 ab	568 b	720 bc
Not infested/sprinkler	402 ab	899 a	836 ab
Not infested/drip	548 a	769 a	876 a

^zMarketable yield was measured on the four plants after thinning from January to June at Oxnard and from April to September at Watsonville. Within a column, means followed by the same letter were not significantly different for a one-way analysis ($\alpha = 0.05$; Tukey-Kramer honestly significant difference).

Table 4. Effect of infestation of strawberry mother plants with *Colletotrichum acutatum* conidia and sprinkler vs. drip irrigation in the nursery on the incidence of *C. acutatum* infection in crowns of daughter plants before and after fruit production.^z

Treatment of mothers with <i>C. acutatum</i> at planting	Irrigation during daughter production	Daughters infected with <i>C. acutatum</i> (% by bioassay)			Daughters infected with <i>C. acutatum</i> in crown (% by quantitative PCR) ^v	
		At harvest at the nursery (%) ^y		7 weeks after transplanting, on either crowns or roots (%) ^{x,w}	7 weeks after ransplanting ^x	After fruit production ^u
		Either crowns or roots	Both crowns and roots			
Infested	Sprinkler	82 a	51 a	100 a	95 a	92 a
Infested	Drip	78 a	29 b	83 a	86 a	70 a
Not infested	Sprinkler	2 b	0 v	8 b	8 b	42 b
Not infested	Drip	0 b	0 v	0 b	0 b	17 b

^zAt planting in the nursery in 2008, mother plants were either dipped into a suspension of *Colletotrichum acutatum* conidia or not and either propagated under drip or sprinkler irrigation. Plants had no signs of anthracnose in the nursery. After harvest at the nursery, daughters suitable for transplantation (crowns with diameter from 7.2–10.9 mm) were selected and either assayed or transplanted into a fruiting field with a completely randomized block design with 12 replicate blocks, each with eight plants (four plants on each of two diagonal rows) per block. Means followed by the same letter are not different ($\alpha = 0.05$). Data were analyzed by Tukey's honestly significant difference except when indicated otherwise.

^yThe percentage infected was determined by a bioassay of 15 plants from each of three replicate bins.

^xAfter 7 weeks in the fruiting field, plants were thinned to four plants per replicate. One thinned plant per replicate was arbitrarily assigned for bioassay and two for quantitative polymerase chain reaction.

^wData were analyzed by contingency analysis. Pairwise comparisons were made by Fisher's exact test using the Bonferroni adjustment for six comparisons ($P = 0.0083$ for $\alpha = 0.05$).

^v*C. acutatum* DNA in crowns was determined by real-time polymerase chain reaction (PCR) with Taqman technology.

^uAll four plants per replicate from each infested treatment were analyzed. One plant per replicate from each non-infested treatment was randomly selected and analyzed.

Table 5. Effect of infestation of strawberry mother plants with *Colletotrichum acutatum* conidia and sprinkler vs. drip irrigation in the nursery on the quantity of *C. acutatum* in crowns of daughter plants before and after fruit production.^z

Treatment of mothers with <i>C. acutatum</i> at planting	Irrigation during daughter production	ng <i>C. acutatum</i> DNA per g dry wt crown tissue ^v			
		All plants		Infected plants only	
		7 weeks after transplanting ^x	After fruit production ^w	7 weeks after transplanting ^x	After fruit production ^w
Infested	Sprinkler	8 a	50 a	16 a	62 a
Infested	Drip	0.7 b	6 b	2 b	17 ab
Not infested	Sprinkler	0.04 b	0.3 c	0.7 b	0.9 b
Not infested	Drip	0.00 b	0.1 c	NA ^y	0.7 b

^zAt planting in the nursery in 2008, mother plants were either dipped into a suspension of *Colletotrichum acutatum* conidia or not and either propagated under drip or sprinkler irrigation. Plants had no signs of anthracnose in the nursery. After harvest at the nursery, daughters suitable for transplantation (crowns with diameter from 7.2–10.9 mm) were selected and either assayed or transplanted into a fruiting field with a completely randomized block design with 12 replicate blocks, each with eight plants (four plants on each of two diagonal rows) per block. Within a column, means followed by the same letter were not significantly different by Tukey's honestly significant difference, $\alpha = 0.05$.

^v*C. acutatum* DNA in crowns was determined by real-time polymerase chain reaction with Taqman technology. Data were analyzed as the log (1 + ng *C. acutatum* DNA) per gram crown tissue. Back-transformed values are shown.

^wAfter seven weeks in the fruiting field, plants were thinned to four plants per replicate. Of the thinned plants, one and two plants from the noninfested and infested mother plant treatments, respectively, per replicate were assayed.

^xAll four plants per replicate from each infested treatment was analyzed. One plant per replicate from each non-infested treatment was randomly selected and analyzed.

^yNot applicable; no infected plants were detected.

The potential for drip to reduce disease risk could result in less fungicide use. Use of drip irrigation in the nursery may also help minimize the negative effects of other water-dispersed pathogens as we have demonstrated here for *C. acutatum* on strawberry.

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